



- [GOAL Events](#)
- [Advocate Magazine](#)
- [Aquademia Podcast](#)
- [Blog](#)
- [Contact](#)
-
-
-
-
-
- [Log In](#)



- [About](#)
 - [Who We Are](#)
 - [Our History](#)
 - [Our Team](#)
 - [Sustainable Development Goals](#)
 - [Careers](#)
- [Membership](#)
 - [Overview](#)
 - [Our Members](#)
 - [Corporate Membership](#)
- [Resources](#)
- [Certification](#)
 - [Best Aquaculture Practices](#)
 - [Best Seafood Practices](#)



[Log In](#)

- [About](#)
 - [Who We Are](#)
 - [Our History](#)
 - [Our Team](#)
 - [Sustainable Development Goals](#)
 - [Careers](#)
- [Membership](#)
 - [Overview](#)
 - [Our Members](#)
 - [Corporate Membership](#)
- [Resources](#)
- [Certification](#)
 - [Best Aquaculture Practices](#)
 - [Best Seafood Practices](#)
- [GOAL Events](#)
- [Advocate Magazine](#)
- [Aquademia Podcast](#)
- [Blog](#)
- [Contact](#)



Responsibility
Responsibility

Toxicity of chemical substances in aquaculture



29 April 2019 Claude E. Boyd, Ph.D.



Difficulties in assessing, establishing safe concentration limits for toxic metabolites



Toxicity in aquaculture systems usually refers to the harmful effects of elevated concentrations of metabolites (carbon dioxide, ammonia, nitrite, and hydrogen sulfide), algal toxins, heavy metals, and agricultural and industrial chemicals. Photo by Darryl Jory.

The dictionary definition of toxicity is “the quality, relative degree, or specific degree of a substance being toxic” and toxic is further defined as a substance containing a material capable of causing injury or death to living things (being poisonous). Stress, injury or death to aquatic organisms resulting from adverse water temperature, low dissolved oxygen concentration, gas supersaturation and suboptimal salinity usually are not referred to by aquaculturists as toxic effects. In aquaculture, toxicity usually is reserved for harmful effects of elevated concentrations of metabolites (carbon dioxide, ammonia, nitrite and hydrogen sulfide), algal toxins, heavy metals and agricultural and industrial chemicals.

Toxicity may be expressed by several reactions of organisms to harmful substances. By slowly and methodically increasing concentrations of potentially toxic substances, a series of responses can be observed. Initially, concentration is too low to evoke a measurable or observable response. At a higher concentration, animals will exhibit behavioral changes such as avoidance, increased respiration, coughing (fish have a response similar to coughing to rid gills of foreign substances), changes in swimming patterns, etc. Increasing concentration of the substance further will cause observable lesions on gills or other body parts, mucus coating on gills and internal lesions. At a greater toxin concentration, animals will exhibit erratic behavior such as swimming at the surface and loss of equilibrium.

Assessing lethal concentrations

Although aquatic animals exhibit symptoms of poisoning or toxicity at lower than lethal concentrations of a substance, the typical endpoint in tests to assess toxicity of a substance is death. In a toxicity test, fish or other aquatic organisms are exposed to a range of toxin concentrations. Mortality at each toxin concentration is recorded after a certain period and adjusted for mortality of animals held under identical conditions in a control (no toxin). Toxicity tests may be run for six to 24 hours, several days, or even for the life cycle of the test organism. Toxicity tests usually are conducted for 24 to 96 hours, and the 96-hour toxicity test is likely the most common.

Toxicity may be reported in various ways. The highest concentration causing no mortality may be reported, or alternatively, the concentration killing a given percentage of the test organisms reported. Most commonly, the concentration that killed 50 percent of test animals is presented (Fig. 1). This concentration is known as the lethal concentration to 50 percent of test animals or the LC50 at a particular exposure time, i.e., 24-hr LC50, 48-hr LC50, 72-hr LC50, or 96-hr LC50.

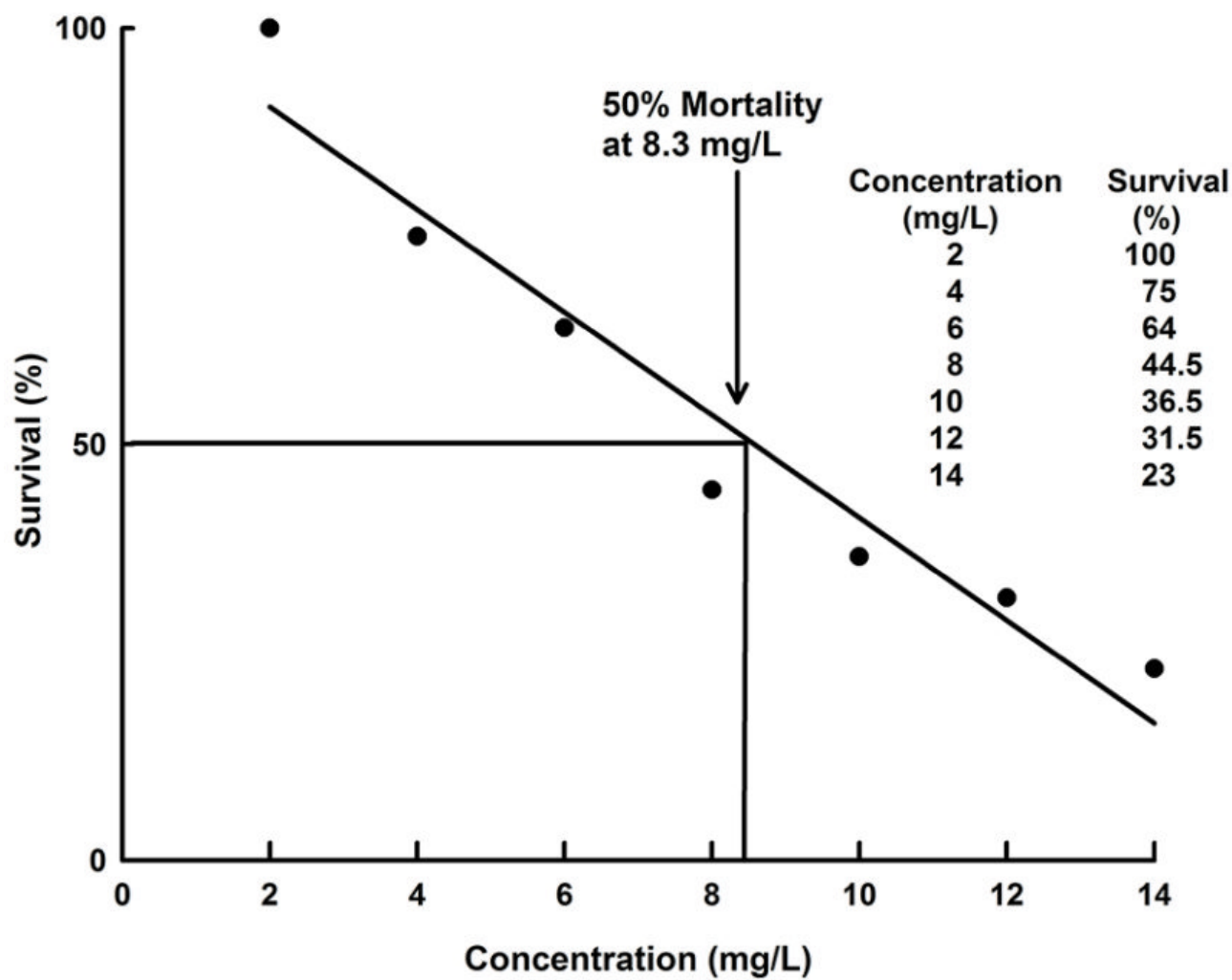


Fig. 1: Graphical estimation of the LC50.

There are difficulties in assessing LC50 with respect to effects of a toxin in natural waters or in aquaculture. Obviously, it is desirable not to have either low level mortality or sublethal effects that stress animals and impair normal behavioral and physiological responses. In aquaculture, stress leads to poor appetite and it weakens immune responses increasing susceptibility to disease. The LC50 does not provide this information directly.

Some investigators have run toxicity tests at low enough concentrations to determine the lowest concentration a toxin necessary to cause mortality. Others have used a sublethal but measurable or observable response such as growth rate, lesions, or behavioral change. Such tests are difficult, time consuming, and expensive to conduct, and few have been conducted in comparison with the number of short-term LC50 tests that have been published.

Results from long-term LC50 tests often are used to extrapolate the expected maximum concentration of a toxin permissible without causing any observable effect to a particular species upon continuous exposure. The ratio of the safe or no-effect concentration from long-term toxicity tests to the LC50 concentration (usually the 96-hour LC50) can be used to predict a safe concentration of the toxin. For example, if the long-term, no effect concentration of a toxin is 0.01 mg per liter and the 96-hour LC50 of this toxin is 0.25 mg per liter, the ratio (often called the application factor) is 0.04. An estimate of the safe concentration of the toxin can be made by multiplying 0.04 by the 96-hour LC50.

Interpreting toxicity test results

The problem with the application factor is that it is not available for many species. Available data often are applied beyond the species for which they were obtained. The application factor also may vary among toxins. Application factors of 0.05 and 0.01 are probably most commonly used. A larger application factor likely should be used for less toxic substances and a smaller one for more toxic substances. Use of the application factor may result in poor decisions in some instances, but it is often the only procedure for use in practical predictions of acceptable toxin concentrations.

Several other issues are related to interpretation of toxicity test results. Toxicity tests are typically run at constant temperature in water of a particular chemical composition, and with constant pH, near saturation with dissolved oxygen and fixed concentrations of the toxin. Many toxins increase or decrease in toxicity with respect to water quality, as will be illustrated for a few examples. Copper and zinc are less toxic in water of high calcium concentration, because calcium ion interferes with metal uptake. In freshwater, chloride ion lessens nitrite uptake across gills of fish through competition for absorption sites. The proportion of toxic, un-ionized ammonia in water increases with total ammonia nitrogen, temperature and pH. Hydrogen sulfide

concentration also decreases in relation to total sulfide concentration as pH increases. Elevated carbon dioxide concentration is more likely to affect aquatic animals when dissolved oxygen concentration is low, because it lessens the ability of hemoglobin to combine with oxygen.

In aquaculture systems, concentrations of pH, temperature, dissolved oxygen and carbon dioxide fluctuate on a daily basis. Toxin concentration also is continually changing because of differential rates of release into the water and loss from the water by detoxification or other means. Exposure time to high concentrations of potential toxins such as carbon dioxide, ammonia, nitrite, hydrogen sulfide, algal toxins, metals, etc., are continuously changing. There have been few studies showing how such fluctuations effect the toxicity of substances to aquaculture species.

When a mortality problem or slow growth by a species cannot be explained by the usual causes, managers have complete water quality analyses conducted. There is a tendency to suspect the cause to be the variable or variables that the analysis report indicates to be outside what the manager considers the normal range.

The decision on presence of toxic concentrations of substances can sometimes be improved by examination of typical LC50 values such as presented in Table 1 for toxic metabolites and aid of an application factor. Nevertheless, it often is virtually impossible to be sure that a problem is associated with a particular variable. This is especially true for algal toxins. Some laboratories have the capability to measure algal toxins, but there is little information on toxic concentrations. With toxic pollutants from agricultural and industrial chemicals, knowledge that one or more such chemicals entered the pond and failure to find evidence of the presence of another toxin often is a strong indicator that the pollutants caused the observed mortality.

Boyd, Toxicity, Table 1

	96-hour LC50 (mg/L)	Estimated safe concentration for extended exposure (mg/L)
Carbon dioxide, freshwater fish	–	20
Carbon dioxide, marine shrimp	60	6
Ammonia, warmwater fish	0.74 – 2.88	0.05 – 0.15
Ammonia, coldwater fish	0.32 – 0.93	0.015 – 0.045
Ammonia, marine fish	0.64 – 1.72	0.05 – 0.15
Ammonia, marine shrimp	0.69 – 2.95	0.05 – 0.15
Nitrite, warmwater fish	7.1 – 140	0.5 – 2.5
Nitrite, coldwater fish	0.24 – 11	0.01 – 0.5
Nitrite, marine fish	–	5 – 50
Nitrite, marine shrimp	9 – 980	0.5 – 15
Hydrogen sulfide, freshwater species	0.02 – 0.05	0.002
Hydrogen sulfide, marine species	0.05 – 0.5	0.005

Table 1. Toxicity information on potentially-toxic metabolites in aquaculture systems.

There also is wide variation in the 96-hour LC50 values for individual metals and agricultural and industrial chemicals, and great variation in toxicity has been found among these chemicals. The 96-hour LC50s for some representative metals were given as follows: manganese, 16 to 2,400 mg per liter; zinc, 0.43 to 9.2 mg per liter; copper, 0.033 to 1.9 mg per liter. The 96-hour LC50s reported for different agricultural and industrial chemicals range from less than 1 mg per liter to over 100,000 mg per liter.

Perspectives

In summary, assigning the cause of an observed sublethal or lethal toxic effect to a particular concentration of a known toxin is not an easy task. Moreover, it is equally difficult to establish safe concentration limits for toxic metabolites in aquaculture systems for use in assessing measured concentrations.

Now that you've finished reading the article ...

... we hope you'll consider supporting our mission to document the evolution of the global aquaculture industry and share our vast network of contributors' expansive knowledge every week.

By becoming a Global Seafood Alliance member, you're ensuring that all of the pre-competitive work we do through member benefits, resources and events can continue. Individual membership costs just \$50 a year.

Not a GSA member? Join us.

[Support GSA and Become a Member](#)

Author



Claude E. Boyd, Ph.D.

School of Fisheries, Aquaculture and Aquatic Sciences
Auburn University
Auburn, Alabama 36849 USA

[117,100,101,46,110,114,117,98,117,97,64,49,101,99,100,121,111,98]

Share

- [✉ Share via Email](#)
- [🐦 Share on Twitter](#)
- [f Share on Facebook](#)
- [in Share on LinkedIn](#)

Tagged With

[aquaculture](#) [LC50](#) [Claude E. Boyd](#) [Toxicity](#) [chemicals](#) [lethal concentration to 50 percent of test animals](#)

Related Posts

[Responsibility](#)

[Calculating chemical treatments for aquaculture production](#)

In calculating chemical treatments to apply to ponds, one must adjust for the percentage of the active ingredients in products. Adjustments for both active ingredient content and specific gravity must be made for liquid products.

[Responsibility](#)

[Calcium and magnesium use in aquaculture](#)

Aquatic plants and animals get the essential nutrients calcium and magnesium from water and food. Calcium concentrations impact the hydration and development of eggs in a hatchery, where calcium carbonate precipitation can be troublesome.

[Responsibility](#)

[Ammonia nitrogen dynamics in aquaculture](#)

The major sources of ammonia in aquaculture ponds are fertilizers and feeds, and problems with high ammonia are most common in feed-based aquaculture.

[Responsibility](#)

[Drying, liming, other treatments disinfect pond bottoms](#)

The traditional way of destroying organisms in pond bottoms is thorough dry-out for a week or longer. Fish toxicants and liming can kill unwanted parasites.

About The Advocate

The Responsible Seafood Advocate supports the Global Seafood Alliance's (GSA) mission to advance responsible seafood practices through education, advocacy and third-party assurances.

[Learn More](#)

Search Responsible Seafood Advocate

Search 

Advertising Opportunities

[2022 Media & Events Kit](#)

Categories

[Aquafeeds](#) > [Health & Welfare](#) > [From Our Sponsors](#) > [Innovation & Investment](#) > [Intelligence](#) > [Responsibility](#) [Responsibility](#) > [Fisheries](#) > [Artículos en Español](#) >

Don't Miss an Article

Featured

- [Health & Welfare](#) [An update on vibriosis, the major bacterial disease shrimp farmers face](#)
- [Uncategorized](#) [A seat at the table: Fed By Blue team says aquaculture needs a stronger voice](#)
- [Responsibility](#) [Quantifying habitat provisioning at macroalgae cultivation locations](#)

Popular Tags

Recent

- [Fisheries](#) [Second Test: Another filler for the fisheries category](#)
- [Fisheries](#) [Test: This is filler for the fisheries Category](#)
- [Aquafeeds](#) [Test Article](#)
- [Responsibility](#) [Study: Climate change will shuffle marine ecosystems in unexpected ways as ocean temperature warms](#)
- [Health & Welfare](#) [Indian shrimp researchers earn a patent for WSSV diagnostic tool](#)



- [About](#)
- [Membership](#)
- [Resources](#)
- [Best Aquaculture Practices \(BAP\)](#)
- [Best Seafood Practices \(BSP\)](#)
- [GOAL Events](#)
- [Advocate Magazine](#)
- [Aquademia Podcast](#)
- [Blog](#)
- [Contact](#)

Stay up to date with GSA

- 
- 
- 
- 
- 

Copyright © 2024 Global Seafood Alliance
All rights reserved.

[Privacy](#)

[Terms of Use](#)

[Glossary](#)