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Genomic IHHN-related DNA sequences in black tiger shrimp



2 September 2013 J.A. Brock, DVM Nathene Antonio Brad Argue, Ph.D.



DNA sequences reacted with OIE-listed IHHN virus diagnostic primers

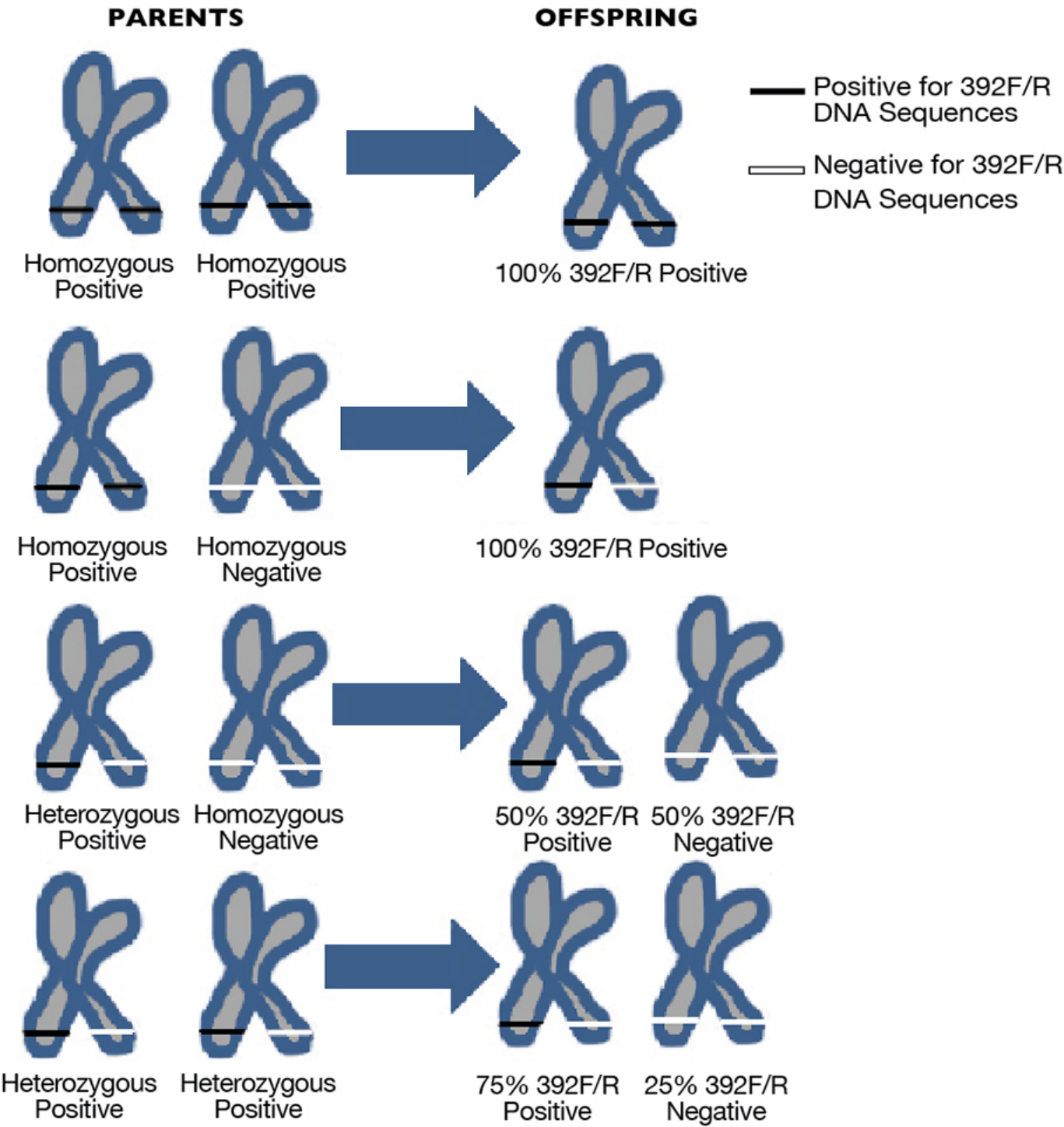


Fig. 1: Examples of possible monodon parent 329F/R IHHN-related genomic sequences and the expected distribution pattern for 392F/R PCR-positive reactors within their offspring.

Infectious hypodermal and hematopoietic necrosis virus (IHHNV) is one of the eight reportable crustacean pathogens listed in the World Organization for Animal Health (OIE) Aquatic Code. In some countries, exclusion and regulatory action are applied if polymerase chain reaction (PCR) tests are positive for IHHNV in samples of farmed or imported live shrimp.

In a 2006 article published in Virus Research, Drs. Kathy Tang and Donald Lightner reported on the occurrence of IHHNV-related DNA sequences within the genome of Penaeus monodon shrimp. The shrimp had been collected from locations in Australia, the Western Indo-Pacific and off the east coast of

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Africa.

In the 2012 online edition of the OIE Aquatic Manual of Diagnostic Tests for Aquatic Animals, the authors stated there are three distinct genotypes recognized for IHNV. Types 1 and 2 are infectious viruses, while Type 3 is non-infectious and recognizable by conventional PCR testing, which detects the IHHN-related DNA sequences present in the genome of *P. monodon*.

The purpose of this article is to provide additional documentation on the geographic range for *P. monodon* where IHHN-related genomic DNA sequences can be expected in PCR tests using recommended primers, as well as provide additional information that demonstrates the sequences are inherited by offspring from their parents according to Mendel’s Law of Segregation.

Diverse distribution

Moana Technologies, LLC’s closed herd of *Penaeus monodon* – housed in the Moana Nucleus Breeding Center (NBC) in Kona, Hawaii, USA – is made up of offspring several generations removed from the original wild, adult founder parents sourced from the South China Sea, Andaman Sea, Bay of Bengal and off the east coast of Africa in the Indian Ocean. IHHN-related genomic sequences are a common feature in Moana *monodon* shrimp.

As an example of a “typical” distribution pattern for IHHN-related genomic sequences in Moana monodon, a listing of PCR test results using a series of IHNV primers is presented in Table 1. The data presented are for a random sample of 20 sub-adult Moana monodon from a grow-out tank in the Moana NBC, a preserved sample of IHNV-infected postlarval *Litopenaeus vannamei* from a hatchery in Asia, and an IHNV-positive control plasmid courtesy of the Aquaculture Pathology Group at the University of Arizona, USA.

Brock, Summary of PCR test results for five IHHN primers, Table 1

Sample	Generation	Sex	SMP-DNA	SYBR/ABP Bactim	IHNV (309F/R)	IHNV (389F/R)	IHNV (392F/R)	IHNV (77012/77353)	IHNV SYBR/ABP
A1	f4	F	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A2	f6	M	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A3	f4	F	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A4	f6	F	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A5	f4	M	Pos.	Pos.	Neg.	Pos.	Pos.	Neg.	Neg.
A6	f5	M	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A7	f6	M	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A8	f5	F	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A9	f5	F	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A10	f6	M	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A11	f6	M	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A12	f6	M	Pos.	Pos.	Neg.	Pos.	Pos.	Neg.	Neg.
A13	f6	F	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A14	f6	M	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
A15	f6	M	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A16	f6	F	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A17	f5	M	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A18	f6	F	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A19	f5	M	Pos.	Pos.	Neg.	Neg.	Pos.	Neg.	Neg.
A20	f6	M	Pos.	Pos.	Neg.	Neg.	Neg.	Neg.	Neg.
B1	L.v.	N.A.	Pos.	Pos.	Neg.	Pos.	Pos.	Pos.	Pos.
IHNV-positive control (plasmid)	IHNV-positive control (plasmid)	N.A.	N.A.	Pos.	Pos.	Pos.	Pos.	Pos.	Pos.

Table 1. Summary of PCR test results for five IHHN primers (309F/R, 389F/R, 392F/R, 77012/77353, Aquabounty IHNV) and two quality-control DNA primers (SMP-DNA and Aquabounty Bactim). Tissue samples from 20 sub-adult *Penaeus monodon* (A1-A20), an IHNV-positive sample of *Litopenaeus vannamei* (B1) and an IHNV-positive control (plasmid).

The data from 2009 show the PCR reaction status for seven conventional primers. Included were two quality-control primers, SMP-DNA shrimp muscle primer and an in-house DNA primer set, and the SYBR IHNV kit Bactim primers. Four of the five IHNV primers are listed in the OIE Aquatic Manual as suitable for detection of IHNV. The fifth was a commercial IHNV kit primer, SYBR/ABP IHNV kit.

Note that for the Moana monodon samples, 50 percent of the shrimp were reactors to the 392F/R primers, 10 percent were PCR positive with the 389F/R primers, and none were PCR positive for the 309F/R, 77012/77353 or SYBR/ABP kit primers for IHNV. Typically, the authors have found that individual shrimp that are PCR positive with the 389F/R primers are also positive with the 392F/R primers. All of the 20 Moana monodon tissue samples were positive for both of the quality-control primers, which indicated the quality of the extracted DNA was suitable for the intended PCR test purpose.

On the other hand, note that the single sample of postlarval *L. vannamei* was PCR positive for the five IHNV primers applied, as was the IHNV plasmid positive control sample. The *L. vannamei* sample was also positive for the two shrimp DNA quality-control primers, which target conservative

DNA sequences in the genome of penaeid shrimp.

Inherited DNA

These PCR test data demonstrated that Moana monodon offspring derived from founder shrimp collected from the Indo-Pacific Region had genomic IHHN-related DNA sequences that reacted with the OIE-listed IHNV diagnostic 392F/R and 389F/R primers. Therefore, caution is advised for the interpretation of conventional PCR test results for IHNV when these primers are applied either singly or in tandem for the detection of IHNV-infected *P. monodon*.

Also note the Table 1 data for the *L. vannamei* sample, which showed the expected PCR test reaction profile from a sample where IHNV was present in the tissue. Note the positive PCR test results for all of the IHNV PCR primers. This example showed the difference between shrimp tissue with IHNV present and shrimp tissue with no virus, as demonstrated with the Moana monodon PCR test results.

The test outcome patterns were clear that where IHNV was not present, the complete viral genome was not present. If the viral genome was not present, the virus itself was not present. Caution is advised in the interpretation of PCR test outcomes using primers that are known to occur naturally within the genome of monodon shrimp.

DNA inheritance

In order to pursue a hypothesis that the sequences were genomic in the monodon shrimp and not related to the presence of actual IHNV, the authors carried out a series of studies to assess the inheritance pattern of the 392F/R DNA in offspring postlarvae spawned from individual male and female parent shrimp.

Individual postlarvae spawned from parents for which the 392F/R IHHN-related DNA sequences had been determined by PCR assay were sampled and tested. The hypothesis was that the 392F/R IHHN-related DNA sequences that occurred within at least one parent would be inherited in offspring shrimp, and the pattern of distribution in the offspring would be in accordance with Mendel’s Law of Segregation (Fig. 1).

Presented in Table 2 are the PCR test outcomes for three separate spawns from Moana breeders. The data show that the 392F/R IHHN-related sequences were inherited according to the Law of Segregation, and the DNA reacting with the PCR test primers must therefore have been within the genome of the parents. These DNA sequences were passed generationally from the parent to the offspring.

Brock, Examples demonstrate how 392F/R IHHN-related DNA sequences pass from parent to offspring, Table 2

Example 1	Example 1	Example 1	Example 1	Example 1	Example 1	Example 1	Example 1	Example 1	Example 1
Parents of P1 Family f6-xxx	SMP-DNA	309F/R	389F/R	392F/R	Chi square statistic for f6-xxx. n = 43	Chi square statistic for f6-xxx. n = 43	Chi square statistic for f6-xxx. n = 43		
Female	Pos.	Neg.	Neg.	Neg.	Category	Observed	Expected	% Expected = 0.581	Chi square value
Male	Pos.	Neg.	Neg.	Pos.	Positive	24	21.5	50	P = 0.45
					Negative	19	21.5	50	
Example 2	Example 2	Example 2	Example 2	Example 2	Example 2	Example 2	Example 2	Example 2	Example 2
Parents of P1 Family f5-xxx	SMP-DNA	309F/R	389F/R	392F/R	Chi square statistic for f5-xxx. n = 50	Chi square statistic for f5-xxx. n = 50	Chi square statistic for f5-xxx. n = 50		
Female	Pos.	Neg.	Neg.	Pos.	Category	Observed	Expected	% Expected = 0.286	Chi square value
Male	Pos.	Neg.	Neg.	Pos.	Positive	49	50.0	100	P = N.A.
					Negative	1	0	0	
Example 3	Example 3	Example 3	Example 3	Example 3	Example 3	Example 3	Example 3	Example 3	Example 3
Parents of P1 Family f5-xxx	SMP-DNA	309F/R	389F/R	392F/R					
Female	Pos.	Neg.	Neg.	Pos.	Category	Observed	Expected	% Expected = 0.286	Chi square value
Male	Pos.	Neg.	Neg.	Pos.	Positive	33	31.5	75	P = 0.593
					Negative	9	10.5	25	

Table 2. Examples demonstrate how 392F/R IHHN-related DNA sequences pass from parent to offspring according to Mendel’s Law of Segregation.

Sequencing analysis Moana has submitted samples of purified DNA amplified by 392F/R primers and excised from agarose gels for sequencing analysis to the Pacific Biosciences Research Center Biotech Core at the University of Hawaii at Manoa. Sequencing results received by e-mail are then further

analyzed using the Basic Local Alignment Search Tool (BLAST).

The samples selected were from representative individual shrimp for the four groupings of genetic stock in the Moana monodon breeding herd. Tissue samples collected from 16 individual monodon shrimp and 16 different families were used to develop the findings presented in Table 3.

Brock, NCBI submission descriptions for IHNV-related genomic sequences, Table 3

NCBI Blastn Accession Number	Description
EU675312.1	IHHNV isolate Au2005
EU675313.1	IHHNV isolate Weipa (Northern Australia), 911 NS2 protein gene
AY124937.1	IHHNV isolate from East Africa, non-structural protein 1 and protein genes
AY362547.1	IHHNV from Thailand, non-structural proteins 1 and 2
AY355307.1	IHHNV isolate from Taiwan
AY590120.1	IHHNV from Australia, non-structural protein 1 and 2 genes

Table 3. NCBI submission descriptions for IHNV-related genomic sequences identified in Moana monodon tissue samples.

Note that for the Moana monodon with IHNV-related genomic DNA sequences, the Blastn accession number results were different from those of the tissue sample that had infectious IHNV (Table 4). The sequence analysis findings further supported the conclusion that the DNA tested was very different between the monodon genomic DNA and the IHNV DNA.

Brock, Submission descriptions for IHNV sequences, Table 4

NCBI Blastn Accession Number	Description, Origin
AF218266.2	IHHNV complete genome
EF633688	IHHNV isolate, Fujian
AY355308.1	IHHNV isolate, Taiwan C
AY355306.1	IHHNV isolate, Taiwan A
AY362548.1	IHHNV from Ecuador, non-structural protein
AF273215.1	IHHNV non-structural proteins 1 and 2
AY590121.1	IHHNV, New Caledonia

Table 4. Submission descriptions for IHNV sequences identified in a sample of preserved *L. vannamei* postlarvae from Asia.

Applications

In 2013, the presence of IHNV-related sequences in the genome of *P. monodon* has been public knowledge for the past seven years. Nevertheless, the authors believe that ongoing discussion about the IHNV-related DNA sequences in the genome of *P. monodon* can help keep people better informed about this irregularity and accept that a PCR-positive test using the 392F/R and 389F/R primers to assay for IHNV does not by itself mean the shrimp were infected with IHNV. As advised in the OIE Aquatic Manual, additional laboratory work is needed to clarify the source of the DNA that reacted positively in the PCR tests.

Moreover, the authors believe that the source location for the DNA sequences for a 392F/R positive PCR test reaction should not be based upon the application of the 389F/R PCR test, because their findings indicated that individual monodon shrimp can have genomic DNA sequences that prime up with these two PCR primer sets.


A second point is that when IHNV is present in shrimp, all PCR primers typically react and display positive test results with the extracted DNA from the sample. This implies that when infectious IHNV is present, DNA for the virus is also present.

However, when infectious IHNV is not present, but shrimp have genomic DNA with the related sequences, positive PCR outcomes will not be found for the five PCR primer sets used. The simple explanation is that no “live” or infectious virus is present, because the virus genome is absent in the sample, and the entire virus genome must be present for an infectious and replicating virus to be present.

A third point is that there may be errors in the published literature regarding the classification of the different genomic types of infectious IHNV, because the analysis work was carried out using tissue samples collected from *P. monodon* prior to learning that the presence of IHNV-related DNA sequences were part of the genomic DNA of *P. monodon*. Thus, if two sources (virus and host) contributed to the extracted DNA that made up the analyzed samples, it would be unreliable to assume the origin of the DNA for the sequence generated in the PCR test was only the IHNV.

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