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# Genetic diversity of HPV parvovirus

Responsible Seafood Advocate logo

1 July 2008 Kathy F.J. Tang, Ph.D. Carlos R. Pantoja, Ph.D. Donald V. Lightner, Ph.D.



# Virus examined at University of Arizona Aquaculture Pathology Laboratory



The hepatopancreatic cells of HPV-infected shrimp exhibit intranuclear inclusion bodies (marked by arrows).

Hepatopancreatic parvovirus (HPV) is a small icosahedral, nonenveloped virus containing a single-stranded linear DNA genome that is placed as a member of (*Parvorividae*). There is no specific gross sign for HPV-infected shrimp, but histology reveals the presence of intranuclear inclusion bodies in hepatopancreatic tubule epithelial cells

Discovered in cultured (*Penaeus merguiensis*) in Singapore in 1982, the virus has been associated with poor growth and mortalities up to 100 percent in larval and postlarval penaeids. It has since been detected in (*P. chinensis*), (*P. semisulcatus*), (*P. monodon*), (*P. stylirostris*) and (*P. vannamei*). HPV has been found in both cultured and wild stocks in Southeast Asia, the Middle East, the Americas, Oceania and Africa.

# **HPV** variation

The authors' laboratory at the University of Arizona maintains an archive of samples of shrimp infected with HPV received from around the world. This has afforded the opportunity to study genetic variation among isolates collected over a wide geographic area.

In recent work, the nucleotide sequences of HPV isolates from Madagascar and Tanzania were compared with HPV sequences deposited in the GenBank genetic sequence database from Korea, Thailand and Australia. Samples of HPV-infected juvenile *P. monodon* were collected from a farm in Madagascar in 2007. HPV-infected postlarval *P. monodon* from Tanzania were sent from a farm near Dar es Salaam in 2008.

DNA was extracted from the hepantopancreas tissue and amplified with HPV-specific primers. The purified and sequenced amplicons indicated 5,742 and 5,685 base pairs of Madagascar and Tanzania HPV, respectively.

# Sequence analysis

The overall mean distance or percentage of difference in nucleotides among the five isolates was 17 percent, with the largest (21 percent) distance between Thailand and Australia isolates. At 12 percent, Madagascar HPV was closest to the Tanzania isolate.

The amino acid sequences of the open reading frames were also compared among the five isolates. ORF is a DNA sequence that could potentially encode a protein. For the left ORF, the overall mean distance was 13 percent. The mid ORF had the lowest (7 percent) distance. The right ORF, which encodes the viral capsid protein, had the highest (24 percent) distance.

In general, the coding sequences for viral structural protein had higher variation than those coding for nonstructural proteins, which are responsible for viral DNA or RNA replication and usually more conserved.

The study revealed a high level of genetic variation among the isolates. HPV appears to have greater genetic diversity than other shrimp viruses such as infectious hypodermal and hematopoietic necrosis virus (IHHNV) and Taura syndrome virus (TSV), which may reflect the fact that it is found over a much wider geographic area than the others.

# **Phylogenetic analysis**

The amino acid sequences of the right ORF were used to construct a phylogenetic tree. Seven isolates representing three genotypes were included in the analysis: type 1 (Korean, Madagascar, Tanzania), type 2 (Thailand and Indonesia) and type 3 (Australia, and New Caledonia).

The two Africa isolates and that from Korea were grouped with a bootstrap value of 75 percent, which is not strong and should be considered tentative until more isolates are analyzed. The isolates from New Caledonia and Australia clustered into one group. HPV from Indonesia and Thailand are closely

related and were clustered together. The isolate clusters reflect geographic distribution of the samples.



Related HPV isolates were clustered within three genotype groups.

## **Impacts of HPV infection**

Although HPV may be highly infectious in wild populations, its effects on shrimp growth are not clear. A 95 percent prevalence of HPV was found in wild populations of *P. merguinesis* in New Caledonia, and *P. monodon* in Madagascar had 90 percent prevalence. This suggested that HPV is very transmissible in the wild, but reports on the impacts of this virus on farmed shrimp are conflicting.

HPV has been linked to growth reduction in farmed *P. monodon* in Thailand, but in Madagascar, HPV infection appears to have no negative effect on shrimp growth. The various effects reported from different areas may be related to differences among viral genotypes, host populations and/or farming practices. For example, the Thailand isolate differs from Madagascar isolate by 20 percent in nucleotide sequence and was shown to be a different genotype. Populations of *P. monodon* in the Indian Ocean are genetically different from those of the western Pacific Ocean.

Also, high stocking density is known to result in reduced growth due to competition for space and food. In Madagascar, the shrimp were cultured at a stocking density of 10-15 per square meter, while those in Thailand are generally stocked at over 75 per square meter. Therefore, the stunted growth reported in HPV-infected *P. monodon* cultured in Thailand may be related to the high stocking density.

## **Madagascar HPV**

African stocks of *P. monodon* are often used as broodstock in other areas of the world because they are free of the major infectious shrimp viruses: white spot syndrome virus, TSV, IHHNV, yellow head virus and infectious myonecrosis virus. However, in Madagascar and Mozambique, nearly 100 percent of both the wild and farmed populations are infected with HPV.

The effects of Madagascar HPV on shrimp growth need to be clarified through controlled experiments. If Madagascar HPV negatively impacts shrimp growth, then stocks need to be monitored for HPV prior to shrimp being stocked in grow-out ponds or exported as broodstock.

(Editor's Note: This article was originally published in the July/August 2008 print edition of the Global Aquaculture Advocate.)

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